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(54) Title: PREPARATION OF FROZEN FERMENTED FOODS USING ANTIFREEZE POLYPEPTIDE-EXPRESSING MICROORGANISMS

## (57) Abstract

Methods and compositions for preparation of frozen fermented food products using antifreeze polypeptide-expressing microorganisms are provided. In particular the invention provides for use of fish antifreeze polypeptide-expressing microorganisms in fermentation of milk to produce and store frozen yogurt.

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PREPARATION OF FROZEN FERMENTED FOODS USING ANTIFREEZE  
POLYPEPTIDE-EXPRESSING MICROORGANISMS

FIELD OF THE INVENTION

The present invention relates generally to methods and reagents useful in  
5 maintaining the quality of frozen food products during frozen storage, particularly  
enhanced storage life and the maintenance of consumer accepted quality of dairy  
products.

BACKGROUND OF THE INVENTION

Refrigeration, particularly freezing, is a common and preferred means for  
10 storing biological materials. Frozen storage generally arrests or considerably slows the  
deterioration of the biological product.

Frozen or refrigerated foods are now a mainstay of the human diet in  
developed nations. Thus extensive research has and is being carried out by food  
scientists to ensure high quality products for the consumers. This is particularly true  
15 with regard to frozen vegetables and frozen deserts such as ice cream and yogurt.

Frozen deserts such as ice cream or yogurt are generally eaten in the  
frozen state. Thus, the texture of the frozen product as well as its flavor is important to  
consumers. Texture is to a large extent governed by the size of the ice crystals.  
Producers of these frozen deserts have gone to considerable effort and expense to ensure  
20 smooth textured products. However, during frozen storage the ice crystals can grow and  
thus roughen and spoil this texture. The growth of ice crystals during frozen storage is  
known as recrystallization. This problem is particularly common when the frozen  
storage conditions are less than ideal, such as during transportation or storage in modern  
frost-free home freezers. After a relatively short period of time at above-zero  
25 temperatures (*i.e.*, above 0°C), or even at sustained freezing temperatures, frozen foods  
can become less desirable or even unsuitable for human consumption due to the ice  
recrystallization process.

Although manufacturers use a variety of techniques to reduce the damage  
associated with recrystallization success has been limited and significant problems

remain. Thus there is a need for new techniques to reduce or prevent the recrystallization process and improve the characteristics of frozen foods. These techniques and compositions should be inexpensive and completely safe and suitable for human consumption.

5 It has been clearly demonstrated that antifreeze polypeptides (AFP) can effectively inhibit ice recrystallization at low ( $\mu\text{g/ml}$ ) concentrations in aqueous solutions and frozen food products (see, e.g., Knight *et al.*, 1984, *Nature* 308:295-296; Knight *et al.*, 1986, *Cryobiology*, 23:256-262; Knight *et al.*, 1988, *Cryobiology*, 25:55-60; Warren *et al.*, U.S. Patent No. 5,118,792). Warren *et al.*, *supra*, have suggested adding  
10 purified antifreeze polypeptides directly to food products prior to freezing to improve preservation characteristics during frozen storage.

At the present time antifreeze proteins are available for commercial use from two sources; the blood serum from a small number of fish species found in cold water, and recombinant DNA techniques such as those described by (but not restricted  
15 to) Warren *et al.*, *supra*. Other sources of antifreeze proteins, such as transgenic plants and animals, are currently being explored. Regardless of the source, the antifreeze polypeptides must be isolated from the medium in which they are found or produced, and subject to extensive purification. These purification procedures are expensive to the point where the cost of the antifreeze polypeptide additions could exceed the value of the  
20 frozen product. Moreover, the purification protocol may introduce contaminants unsafe for consumption.

In the view of the inherent value to producers and consumers of inhibiting ice recrystallization in frozen fermented dairy products, and the fact that antifreeze polypeptides are very effective in this regard, it is desirable to develop methods to  
25 incorporate the antifreeze polypeptides into the food products in the most efficient and cost effective method possible.

An ideal method of incorporating antifreeze polypeptides into frozen fermented food products is to have the organism responsible for the fermentation process produce the antifreeze proteins while fermenting the food. A number of antifreeze  
30 polypeptides and their genes have been well characterized and sequenced (see, e.g. Anathanarayanan, 1989, *Life Chem. Rep.* 7:1-32 and Davies *et al.*, 1990, *FASEB Journal* 4:2460-2467). Several of these genes have been expressed in bacteria, transgenic plants, fish and *Drosophila* (see, e.g., Fletcher *et al.*, 1988, *Can. J. Fish. Aquat. Sci.* 45:352-

In a most preferred embodiment the invention comprises incubating milk with bacterial species *Lactobacillus bulgaricus* and *Streptococcus lactis* that are capable of fermenting milk to produce yogurt and capable of secreting an ocean pout type III antifreeze polypeptide; incubating the bacteria and milk under conditions that produce yogurt; and freezing the yogurt at a temperature below  $-5^{\circ}\text{C}$ , so as to produce frozen yogurt.

The invention also provides a composition comprising yogurt and a microorganism wherein the microorganism comprises a gene encoding a fish antifreeze polypeptide.

10

## DETAILED DESCRIPTION

### Definitions

As used herein, "fermentation" refers to the chemical conversion of carbohydrates or proteins in foods through the use of microorganisms. In this process carbohydrates are often converted to lactic acid.

15

As used herein, "food product" refers to a foodstuff (a substance that can be used, or prepared for use, as food) that can be transformed by the action of a fermenting microorganism to a fermented food product.

As used herein "fermented food product" refers to an edible food prepared by a process that includes fermentation by a microorganism.

20

As used herein "yogurt" refers to a dairy product produced by the lactic acid fermentation of milk by the action of microorganisms.

25

As used herein "Antifreeze polypeptides" (AFPs) refers to macromolecules found in the body fluids of some animals and plants, which have the commonly known property that they reduce non-colligatively the freezing point of water. Antifreeze polypeptides are also known as "thermal hysteresis proteins." As used herein, "antifreeze polypeptides" includes glycoproteins as well as chemically synthesized, and recombinantly produced polypeptides having a protein sequence with substantial similarity to a naturally occurring AFP and retaining the properties of an antifreeze polypeptide.

30

As used herein "fish antifreeze polypeptide" refers to an AFP that is found in nature in a fish, as well as chemically synthesized and recombinantly produced

357; Rancourt *et al.*, 1987, *Mol. Cell. Biol.* 7:2188-2195; Kenward *et al.*, 1993, *Plant Mol. Biol.* 23:377-385; Li *et al.*, 1991, *Protein Engineering* 4:995-1012; and Sønnichsen *et al.*, 1993, *Science* 259:1154-1157).

5 In view of the widespread popularity of frozen fermented dairy products such as yogurt it would be desirable to develop methods for producing such products more efficiently at lower cost and with better flavor and texture. The present invention provides methods for achieving this goal.

### SUMMARY OF THE INVENTION

10 The present invention provides methods for preparing a frozen fermented food product. This method comprises the steps of (a) contacting a food product with a microorganism that is capable of secreting a fish antifreeze polypeptide, wherein the microorganism is capable of fermenting the food product to produce the fermented food product, (b) incubating the food product with the microorganism under conditions in which fermentation takes place so that a fermented food product is produced having the  
15 antifreeze polypeptide present in an amount effective at inhibiting recrystallization of the product; and (c) freezing the fermented food product at a temperature below -5°C, so as to produce a frozen fermented food product.

In one embodiment the food product is a dairy product (*e.g.*, milk) which can be fermented to produce yogurt, buttermilk or cheese.

20 The microorganism of the invention is usually a bacterium (*e.g.*, *Lactobacillus bulgaricus*; *Streptococcus cremoris*, *Streptococcus lactis*; *Bifidobacterium bifidum*, *Bifidobacterium longum*) but may also be a fungus such as a yeast (*e.g.*, *Saccharomyces fragilis*, *Saccharomyces cerevisiae*, *Saccharomyces lactis*, and others). According to the invention these microorganisms are genetically engineered so that they  
25 are capable of secreting a fish antifreeze polypeptide (or a peptide with substantial sequence similarity to a fish antifreeze polypeptide and with antifreeze properties similar to a fish antifreeze polypeptide). In a preferred embodiment, the antifreeze polypeptide is an animal antifreeze polypeptide, with a fish antifreeze polypeptide preferred. Most preferred is microorganism capable of expressing an ocean pout type III antifreeze  
30 polypeptide (*see, e.g.*, Hew *et al.*, 1984, *J. Comp. Physiol.* B155:81-85; Li *et al.*, 1985, *J. Biol. Chem.* 260:12904-12909).

polypeptides having a protein sequence with substantial similarity to a naturally occurring fish AFP and retaining the properties of a antifreeze polypeptides.

As used herein, "recombinantly produced polypeptides" refers to a polypeptide produced using recombinant DNA techniques. Recombinant DNA techniques are well known and are characterized by the joining of at least two segments of DNA that are not naturally joined in nature (e.g., a bacterial promoter and a fish polypeptide coding sequence).

As used herein, "substantial similarity" denotes a characteristic of a polypeptide sequence or nucleic acid sequence, wherein the polypeptide sequence has at least 70 percent sequence identity, preferably 80 percent sequence identity, and most preferably 90% sequence identity compared to a reference sequence (e.g., a naturally occurring antifreeze polypeptide), and the nucleic acid sequence has at least 80 percent sequence identity and preferably 90% sequence identity compared to a reference sequence. The reference sequence may be shorter than the full-length naturally occurring polypeptide or nucleic acid sequence but will be at least 12 residues long for the case of a polypeptide and at least 36 bases long for the case of a nucleic acid.

#### Description

The present invention provides methods for preparing a frozen fermented food product by adding a microorganism that is capable of fermenting the food product to produce the fermented food product and also is able to secrete a fish antifreeze polypeptide. The use of a microorganism that both secretes an AFP and ferments the food product has several advantages over other methods for affecting ice crystal formation and freezing temperature. For example, the claimed method avoids the costly necessity for purifying an AFP prior to addition to a food product. In addition, this will eliminate any possible contamination from the purification protocol and the pyrogenicity associated with foreign microorganisms. Furthermore, because the AFP is secreted by the fermenting microorganism of the claimed invention, this process requires fewer steps than other methods.

The food product of the invention is usually milk but other foods that are fermented to produce an edible fermented food may also be used. Examples include cabbage (which can be fermented to produce sauerkraut), cucumbers (which can be

fermented to produce pickles) and soybeans (which can be fermented to produce miso and other products).

In one step of the claimed method, the food product is contacted or mixed with a microorganism capable of fermenting the food product. Examples of  
5 microorganisms useful in food fermentation are well known (see, e.g., van de Guchte, 1992, *FEMS Microbiology Reviews*, 88:73-92).

In a preferred embodiment the food product is milk (e.g., from a cow [i.e. bovine], ewe, mare, or goat). The action of fermenting microorganisms, typically  
10 bacteria, on the milk produces yogurt, buttermilk, or certain cheeses, according to the choice of the bacteria and the conditions of incubation. In a most preferred embodiment the method of the invention will be used to produce yogurt from milk. Yogurt is referred to by a variety of names around the world. Table 1 provides a list of the names and country of origin of the common varieties.

Methods for yogurt production can be found in *Functions of Fermented*  
15 *Milk* edited by Nakazawa and Hosono, 1992, published by Elsevier Applied Science, London-New York, p. 32, which is incorporated herein by reference. In the United States yogurt is produced from either whole or skim milk from cows. The milk is standardized to 10.5 to 11.5% solids, heated to above 90°C (30 to 60 minutes) to destroy any contaminating microorganisms, and then cooled. The material is then inoculated  
20 with a mixed culture of *Streptococcus thermophilus* and *Lactobacillus bulgaricus* in a 1:1 ratio. The combined action of these two organisms is usually needed to obtain the desired flavor and acid in the products. In other instances, other high fermenting bacteria including bulgarian bacteria, *L. jugarti*, *L. acidophilus*, *Bifido bacterium*, spp. Yeast and lactic fungi have also been used. Examples of bacteria and other organisms  
25 used for the fermentation of milk to produce yogurt are given in Table 2.

According to the invention, the microorganisms will be genetically engineered (i.e., employing the techniques of recombinant DNA technology) so that they are able to secrete a fish antifreeze polypeptide. Examples of fish antifreeze polypeptides include type I AFP of winter flounder; type II AFP of sea raven, herring and smelt; type  
30 III AFP of ocean pout and wolffish, and AFGP (antifreeze glycoprotein) of cods and nototheniids (see, e.g. Davies et al., 1990 *FASEB Journal* 4:2460-2467, and Ewart et al., 1992, *Biochem. Biophys. Res. Comm.* 185:335-340).

Most preferred is microorganism capable of expressing an ocean pout type III antifreeze polypeptide. The ocean pout type III antifreeze polypeptide is preferred because it has no amino acid bias and has been shown to be active when expressed in *E. coli* (Li *et al.*, 1991, *Protein Engineering* 4:995-1012; Sönnichsen *et al.*, 1993, *Science*, 259:1154-1157). In addition, the type III AFP is preferred because type I AFP of winter flounder may not be stable at the fermentation temperature and type II AFP may not be correctly folded in bacterial system and is very susceptible to reduction.

The methods for engineering bacteria and fungi capable of expressing and secreting a heterologous polypeptide are well established (*see, e.g.*, Maniatis *et al.* (1982), *Molecular Cloning, A Laboratory Manual*, Cold Spring Harbor Press, Cold Spring Harbor, New York; Berger and Kimmel, *Guide to Molecular Cloning Techniques, Methods in Enzymology* 152 (Academic Press, Inc., San Diego, CA); Simon *et al.*, 1986, *Appl. Environ. Microbiol.* 52:394-395; and von Wright *et al.*, 1985, *Appl. Environ. Microbiol.* 50:1100-1102, all of which are incorporated herein by reference)

The production of microorganisms capable of expressing and secreting an AFP can be carried out in a variety of ways that will be apparent to one of ordinary skill. The DNA sequence encoding the AFP will preferably be operably linked (*i.e.*, positioned to ensure the functioning of) to an operon which allows the DNA to be transcribed (into an RNA transcript) and translated into a polypeptide in the microorganism. Promoters for both bacteria and fungi are well known in the art. Preferred operons for expression in lactic acid bacteria include the lactose operon of *S. thermophilus* or lac ABCDFEGX operon of *L. lactis* because they have been used successfully to drive foreign gene expression in the hosts (*see, e.g.*, Simons *et al.*, 1993, *J. Bact.* 175:5186-5175; Mollet *et al.*, 1993, *J. Bact.* 175:4315-4324).

The AFP may be expressed as a fusion polypeptide for increased stability or other beneficial properties. Furthermore the AFP polypeptide may be modified via a modification of the gene encoding the polypeptide. In general, modifications of the genes may be readily accomplished by a variety of well-known techniques, such as site-directed mutagenesis (*see, e.g.*, Gillman and Smith, 1979, *Gene* 8:81-97 and Roberts *et al.*, 1987, *Nature* 328:731-734).

The microorganisms of the invention are capable of secreting the AFP. Accordingly, the AFP will preferably be linked to a signal peptide sequence. Examples of suitable signal peptide sequences include those from the *usp45* gene of *L. lactis ssp*

**Table 2**  
**Microorganisms commonly used in fermented milk and lactic drinks**

	<u>Genus</u>	<u>Habit</u>	<u>Fermentation</u>	<u>Main Species</u>
5	Streptococcus <sup>a</sup>	Coccal chains	Homo	S. cremoris, lactis, thermophilus
	Leuconostoc <sup>b</sup>	Coccal pairs	Hetero	L. citrovorum, mesenteroides
10	Lactobacillus <sup>c</sup>	Rods	Homo	L. acidophilus, bulgaricus, casei, jugurti, lactis
	Bifidobacterium	Rods	Hetero	B. bifidum, breve, longum
15	<b>Others:</b> Yeasts ( <i>Torulopsis holmil</i> ; <i>Saccharomyces fragilis</i> , <i>cerevisiae</i> , <i>lactis</i> ; <i>Candida pseudotropicalis</i> , etc.) Fungi ( <i>Geotrichum candidum</i> ) Acetic acid bacteria ( <i>Acetobacter aceti</i> , <i>rasens</i> )			
20				
	<sup>a</sup> Now <i>Lactococcus lactis</i> subsp. <i>cremoris</i> , <i>Lac, lactis</i> subsp. <i>lactis</i> and <i>S. thermophilus</i> . <sup>b</sup> Now <i>L. mesenteroides</i> subsp. <i>cremoris</i> and <i>L. mesenteroides</i> subsp. <i>mesenteroides</i> . <sup>c</sup> Now <i>L. acidophilus</i> , <i>L. delbrueckii</i> subsp. <i>bulgaricus</i> , <i>L. casei</i> subsp. <i>casei</i> , <i>L. helveticus</i> biovar. <i>jugurti</i> and <i>L. delbrueckii</i> subsp. <i>lactis</i> .			
25				

**Table 1**  
Names used to describe types of yogurt

	<u>Product Name</u>	<u>Country of Origin</u>
5	Jugurt/Eyran/Ayran	Turkey, etc.
	Busa	Turkestan
	Kissel Mleka	Balkans
	Urgotnic	Balkan Mountains
	Leban/Laban	Lebanon/Arab countries
10	Zabady (Zabbady)	Egypt/Sudan
	Mast/Dough	Iran/Afghanistan
	Roba	Iraq
	Dahi/Dadhi/Dahee	India
	Mazun/Matzoon/Matsun/ Matsoni	Armenia
15	Katyk	Transcaucasia
	Tiaourti	Greece
	Cieddu	Italy
	Mezzoradu	Sicily
	Gioddu	Sardinia
20	Biokys	Czechoslovakia
	Karmdinka	Poland
	Tarho	Hungary
	Tykmaelk/Ymer	Hungary
	Villi (Fiili)	Finland
25	Filmjolk/Fillbunke/ Surmelk/Taettemjolk/ Tettemelk	Scandinavia
	Iogurte	Brazil/Portugal
	Proghurt	Chile
	Skyr	Iceland
	Gruzovina	Yugoslavia
30	Kefir/Donskaya/Varentes	Soviet Union
	Kurunga/Koumiss/ Ryazhenka/Guslyanka	
	Tarag	Mongolia
	Shosim/Sho/Thara	Nepal

*lactis* MG 1363 and the *L. lactis* ssp *cremoris* SK11 cell envelop associated protease gene (van Asseldonk *et al.*, 1990, *Gene* 95:155-160; De vos *et al.*, 1989, *J. Dairy Sci.* 72:3398-3405). For bacteria such as *L. lactis* the usp45 signal peptide is preferred since it derives from the same host. In one preferred embodiment the AFP gene is linked to a transcription termination sequence to ensure correct termination of AFP transcription in the host system.

An AFP gene construct including elements described above is constructed using plasmids such as pUC19, pNZ18 and pDBN183 as vectors (Solaiman *et al.*, 1992, *Plasmid*, 28:25-36). The AFP gene construct is incorporated into the genome of a lactic acid bacterial species using homologous recombination techniques (Mollet *et al.*, 1993, *J. Bact.*, 175:4315-4324). The lactic acid bacteria and *E. coli* strains can be maintained as recommended by Maniatis *et al.* in *Molecular Cloning, A Laboratory Manual*, *supra*; and Chagnand *et al.*, 1992, *Can. J. Microbiol.* 38:67-74.

Fish antifreeze-expressing (FAE) microorganisms may be applied to food products in any conventional way. In the case of products such as milk, the bacteria or fungus can be mixed intimately with the foodstuff that is to be fermented and frozen. It will be known by those of skill that mixtures of different microorganisms are sometimes used to produce the desired product. For example, in preparation of yogurt, *S. thermophilus* and *L. bulgaricus* are often used together.

The number of FAE microorganisms added to the food product will depend on the properties of the microorganisms and of the food. Generally, lactic acid FAE starter bacteria ( $10^{10}$  -  $10^{11}$  per ml) are incubated at 1-5% into pasteurized and cooled milk such that the proportion results in an appropriate amount of antifreeze polypeptide in the product. The amount of AFP in the product should be an amount effective at preventing or inhibiting ice recrystallization (1-100 mg/liter milk). This can be determined using the splat-cooling assay described by Knight *et al.* (1988) *Cryobiology*, vol. 25, pp. 55-60.

In another step of the method, the fermented food product is frozen using conventional freezer operations, such as blast freezers (-20° to 40°C) or contact plate freezers (-30° to 40°C) or vacuum freeze driers. It will be apparent to one of ordinary skill that numerous variations of the aforementioned embodiments are possible.

## EXAMPLES

The invention is illustrated by the following examples. These examples are offered by way of illustration, not by way of limitation.

### Example 1

#### Construction of a strain of lactic acid bacteria that produce antifreeze polypeptide - Method I

5 To engineer a lactic acid bacterium that produce antifreeze proteins several steps are involved. The first step is selection and preparation of a chromosomal site for the AFP gene integration. A native operon of a strain of lactic acid bacteria such as the  
10 lactose operon of the *S. thermophilus* or *L. lactis* genome, consisting of the *lacS* (lactose permease) and *lacZ* ( $\beta$ -galactosidase) genes, is used for the integration of an antifreeze protein gene. Integration of an AFP gene into such an operon should preserve its correct function. The AFP gene should become a functional part of the operon and be regulated similarly (see, e.g., Simons *et al.*, 1993, *J. Bact.*, 175:5186-5175, and Mollet *et al.*,  
15 1993, *J. Bact.* 175:4315-4324).

To do this, the *lacS* and *lacZ* genes from the host bacteria are cloned by PCR procedures or conventional gene cloning methods. At least one restriction enzyme site is generated between the two genes by designing particular primer sequences for the  
20 PCR reactions. The restriction enzyme sites are generated for convenient segment linkage and insertion of DNA fragment. An antibiotic resistance marker gene such as an ampicillin or erythromycin resistance gene is inserted into the generated restriction enzyme site. The *lacS*-Amp<sup>r</sup>-*lacZ* DNA in an appropriate vector such as pNZ932 is transformed into the lactic acid bacterial strain (see, e.g. Simons *et al.*, 1993, *J. Bact.* 175:5186-5175). Ampicillin-resistant transformants will be selected, and gene  
25 integration will be verified using PCR and DNA sequencing.

The second step involves construction of an antifreeze protein gene cassette. Using the nucleotide sequence derived from a cloned ocean pout type III AFP gene, an appropriate type III AFP gene is assembled from synthetic oligonucleotides using the preferential codons of the host (see, e.g., Mercenier, 1990, *FEMS Microbiology Reviews*, 87:61-78, and van Asseldonk *et al.*, 1992, *FEMS Microbiology Reviews* 88:73-92). To make a bacteria secrete AFP, a signal peptide sequence (SP)  
30 from homologous genes such as the *usp45* gene of *L. lactis ssp lactis* MG 1363 and the

*L. lactis ssp cremoris* SK11 cell envelop-associated protease genes is fused to the 5'-end of the type III mature AFP coding sequence (van Asseldonk *et al.*, 1990, *Gene* 95:155-160, and De vos *et al.*, 1989, *J. Dairy Sci.* 72:3398-3405). The AFP gene cassette, including a signal peptide sequence and an AFP gene will be inserted between the *lacS* and *lacZ* genes *in vitro* to generate a *lacS*-SP-AFP-*lacZ* construct.

Finally, the *lacS*-SP-AFP-*lacZ* DNA is incorporated into the genome of the ampicillin-resistant bacteria generated from step one by homologous recombination techniques. Gene replacement of *lacS*-Amp<sup>r</sup>-*lacZ* by *lacS*-SP-AFP-*lacZ* is initially selected by their ability to grow on medium in the presence or absence of ampicillin. AFP gene integration is confirmed by inverse PCR, Southern blot analysis and DNA sequencing.

### Example 2

#### Construction of a strain of lactic acid bacteria that produces antifreeze polypeptide - Method II

An alternative method to generate an antifreeze protein-producing lactic acid bacterium is to eliminate the step of integrating an antibiotic resistance gene construct into the genome of the bacterium. In this case, experiments will be carried out as described in steps 2 and 3 in Example 1. A labelled AFP gene fragment will be used to identify recombinant clones, and AFP gene integration will be further confirmed by inverse PCR and DNA sequencing.

### Example 3

#### Use of antifreeze polypeptide expressing bacteria in yogurt production

The genetically engineered bacterium can be directly used to prepare starters for making frozen yogurt. In brief, a small quantity of the genetically engineered stock culture (frozen or cold stored culture) is cultured in 0.5-1 liter of pasteurized skim milk to prepare a mother culture. Intermediate fermentation is then carried out to scale up 10-100 times by stages from the mother culture to the bulk starter which can be added directly to pretreated milk. Alternatively, bulk starter can be prepared directly from frozen stocks. The milk mixture is prewarmed to 55-60°C and then homogenized (150-200 Kg/cm<sup>2</sup>) and pasteurized (90-95°C for 5-10 minutes or 120-130°C for a few seconds), and cooled to a temperature range within which the lactic acid

bacteria will not be damaged (45-48°C). The starter bacteria are applied to inoculate with pretreated homogenized milk at 30-45°C. When the acidity reaches a certain prescribed level such as 0.8% and the amount of AFP in the product reaches required concentration (1-100 mg/liter milk), the fermented milk is cooled to 15-20°C to suppress bacterial activity. These products are used to make soft frozen yogurt, hard frozen yogurt, and mousse yogurt by adding different percentages of fruit syrup, sugar, stabilizers, fruit juice and emulsifiers to a cold fermented milk base.

#### Example 4

##### Production of frozen yogurt

Soft frozen yogurt is made by adding 20% fruit syrup and stabilizers and emulsifiers to 80% of a cold fermented milk base, and then filling into containers with a 50-60% overrun using a normal ice cream freezer. The product is stored at 0-6°C.

Hard frozen yogurt is made of 35% fruit juice, the overrun is 70-80%, and storage temperature is below -25°C. Mousse yogurt is made by mixing a fermented milk base with a warm mousse base (a homogenized mixture of skim milk, sugar, stabilizers and emulsifiers). The product is stored at below 0°C.

The above examples are provided to illustrate the invention but not to limit its scope. Other variants of the invention will be readily apparent to one of ordinary skill in the art and are encompassed by the appended claims. All publications, patents, and patent applications cited herein are hereby incorporated by reference.

**WHAT IS CLAIMED IS:**

1. A method for preparing a frozen fermented food product comprising the steps:
  - a) contacting a food product with a microorganism capable of secreting a fish antifreeze polypeptide, wherein the microorganism is capable of fermenting the food product to produce the fermented food product;
  - b) incubating the food product with the microorganism under conditions in which fermentation takes place so that a fermented food product is produced having the fish antifreeze polypeptide present in an amount effective at preventing or inhibiting ice recrystallization of the product during storage; and
  - c) freezing the fermented food product at a temperature below  $-5^{\circ}\text{C}$ . so as to produce a frozen fermented food product.
2. The method of claim 1 wherein the food product is a dairy product.
3. The method of claim 2 wherein the food product is bovine milk.
4. The method of claim 3 wherein the fermented food product is yogurt.
5. The method of claim 3 wherein the fermented food product is selected from the group consisting of buttermilk and cheese.
6. The method of claim 1 wherein the microorganism is a bacteria.
7. The method of claim 6 wherein the bacteria is selected from the group consisting of *Streptococcus cremoris*, *Streptococcus lactis*, *Streptococcus thermophilus*, *Leuconostoc citrovorum*, *Leuconostoc mesenteroides*, *Lactobacillus acidophilus*, *Lactobacillus bulgaricus*, *Lactobacillus casei*, *Lactobacillus jugurri*, *Lactobacillus lactis*, *Bifidobacterium bifidum*, *Bifidobacterium breve*, and *Bifidobacterium longum*.

8. The method of claim 1 wherein the microorganism is a fungus.
9. The method of claim 8 wherein the fungus is a yeast.
10. The method of claim 9 wherein the yeast is selected from the group comprising *Torulopsis holmil*, *Saccharomyces fragilis*, *Saccharomyces cerevisiae*, *Saccharomyces lactis*, and *Candida pseudotropicalis*.
11. The method of claim 1 wherein the fish antifreeze polypeptide is from an ocean pout.
12. The method of claim 11 wherein the antifreeze polypeptide is a ocean pout type III antifreeze polypeptide.
13. A method for preparing frozen yogurt comprising the steps:
  - a) contacting milk with a microorganism capable of secreting a fish antifreeze polypeptide, wherein the microorganism is capable of fermenting milk to produce yogurt;
  - b) incubating the microorganism and milk under conditions that produce yogurt;
  - c) freezing the yogurt at a temperature below -5° C. so as to produce frozen yogurt.
14. The method of claim 13 wherein the microorganism is selected from the group: *Streptococcus cremoris*, *Streptococcus lactis*, *Streptococcus thermophilus*, *Leuconostoc citrovorum*, *Leuconostoc mesenteroides*, *Lactobacillus acidophilus*, *Lactobacillus bulgaricus*, *Lactobacillus casei*, *Lactobacillus jugurii*, *Lactobacillus lactis*, *Bifidobacterium bifidum*, *Bifidobacterium breve*, *Bifidobacterium longum*, *Torulopsis holmil*, *Saccharomyces fragilis*, *Saccharomyces cerevisiae*, *Saccharomyces lactis*, and *Candida pseudotropicalis*.
15. The method of claim 13 wherein the antifreeze polypeptide is a ocean pout type III antifreeze polypeptide.

16. A composition comprising yogurt and a microorganism wherein the microorganism comprises a gene encoding a fish antifreeze polypeptide.

17. The composition of claim 16 wherein the gene encodes a ocean pout type III antifreeze polypeptide and the microorganism is a bacteria selected from the group comprising *Streptococcus thermophilus* and *Lactobacillus bulgaricus*.

# INTERNATIONAL SEARCH REPORT

Int'l Application No  
PCT/CA 95/00568

**A. CLASSIFICATION OF SUBJECT MATTER**  
IPC 6 A23L1/03 A23C9/123 A23C19/032

According to International Patent Classification (IPC) or to both national classification and IPC

**B. FIELDS SEARCHED**  
Minimum documentation searched (classification system followed by classification symbols)  
IPC 6 A23L A23C A23G

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO,A,94 03617 (UNILEVER) 17 February 1994 see claims 1-13 ---	1-17
A	PATENT ABSTRACTS OF JAPAN vol. 16 no. 13 (C-901) & JP,A,03 232896 (SUMITOMO CHEM CO) 16 October 1991, see abstract ---	1
A	EP,A,0 424 771 (SOCIÉTÉ DES PRODUITS NESTLÉ) 2 May 1991 see claims 1-22 ---	1-17
A	WO,A,90 13571 (DNA PLANT TECHNOLOGY CO) 15 November 1990 see page 21, line 3 - line 17; claims 1-12 --- -/--	1-27

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

### \* Special categories of cited documents :

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- \*T\* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- \*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
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- \*Z\* document member of the same patent family

Date of the actual completion of the international search

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# INTERNATIONAL SEARCH REPORT

International Application No

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## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>EP,A,0 443 543 (SNOW BRAND MILK PRODUCTS)  28 August 1991  see claims 1-7</p> <p>-----</p>	1

# INTERNATIONAL SEARCH REPORT

Information on patent family members

Int'l Application No

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